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Prevalence of Soil Transmitted Helminths infection in domesticated pigs and dogs of Morogoro region, Tanzania

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Abstract

Soil-transmitted helminths (STH) infections are caused by a group of nematode worms that require development in the soil to become infective. These infections cause ill health in animals; some are responsible for zoonotic infection. Zoonotic STH of dogs and pigs include; Ancylostoma species, Toxocara canis, and Ascaris suum. This study was carried out due to limited information on STH infections in domesticated pigs and dogs in Tanzania. The study aimed to determine the prevalence of STH in domestic pigs and dogs in Morogoro Municipality and Mvomero District, Tanzania. This was a cross-sectional study which involved a total of 281 dogs and 237 pigs from two districts in the two study areas. The study animals were randomly selected and faecal samples collected and examined for STH using a simple flotation technique. The data collected was entered in Excel and analysis done using Stata version 14. Prevalence of the STH was computed for both animal species and chi-square used to obtain p-values as the test for statistical significance. The prevalence of STH in dogs was 50.53% with Ancylostoma species being the most prevalent (47.33%), followed by Toxocara (12.46%), Trichuris (2.49%) and Strongyloides species (1.07%). The STH prevalence in pigs was 85.23% and the isolated species were Oesophagostomum (81.01%), Trichuris (12.24) and Ascaris species (5.49%). Males and pigs aged ≥1 year were more infected with STH species with a p-value of 0.025 and 0.016, respectively. Dewormed dogs were observed to be less likely to be infect with STH species (p-value = 0.002). Results of this study show that STH species were prevalent gastrointestinal parasites in dogs and pigs in the study area. Mixed infection was common in both animal species. Measures for control of STH infections in dogs and pigs should be practiced to improve animal health.

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Introduction

Soil-transmitted helminths (STH) are a group of parasitic nematodes that infect humans and other animals and are transmitted through ingestion of infective eggs or larvae, or penetration of infective larvae into the skin from the environment contaminated by faeces from infected animals (Massetti *et al.,* 2020). Among domesticated animals, STHs occur frequently in dogs, cats and pigs.

Dogs are known to play a crucial role in the transmission of STHs to humans and other animals (Crompton, 2000). Most parasitic

nematodes that infect dogs belong to the STH group (Yacob *et al.*, 2007). These nematodes include *Ancylostoma caninum*, *Ancylostoma braziliense*, *Ancylostoma ceylanicum*, *Uncinaria stenocephala*, *Trichuris vulpis*, *Toxascaris leonina* and *Toxocara canis* (Jia-Chi *et al.*, 2016; Traversa, 2012). STH infections in dogs are characterized by bloody diarrhoea, abdominal pain, poor growth, anaemia and death in cases of acute and heavy infections (Agustina *et al.*, 2021; Chidumayo, 2018).

Dogs are also known to harbour STH reported to be zoonotic in different parts of the world. These zoonotic STH include the hookworms, *A. caninum, A. braziliense, A. ceylanicum* and *U. stenocephala* and the ascarid, *Toxocara canis* (Jia-Chi *et al.*, 2016; WHO, 2016). Canine hookworms cause cutaneous larva migrans in humans (Traub *et al.*, 2021). *Ancylostoma braziliense* is the only species capable of causing creeping eruptions and *A. caninum* can lead to aphthous ileitis and eosinophilic enteritis (Walker *et al.*, 1995). In humans, *Toxocara* can cause visceral larva migans, which include ocular toxocariasis and in children, neurological toxocariasis (Strube *et al.*, 2013; Walsh and Haseeb, 2014).

The major STH infecting domestic pigs are Ascaris suum, Trichuris suis, Strongyloides ransomi, Trichostrongylus species and Oesophagostomum species (Roepstorff & Nansen, 1998). Domestic pigs infected with one or more of these helminths could have diarrhoea, decreased litter size and stunted growth while ascariosis caused by A. suum is among the leading cause of liver condemnation resulting from migration of larvae stage two within the liver tissues (Nganga et al., 2008; Tumusiime et al., 2020). Ascaris suum and maybe T. suis are reported to be zoonotic to human beings (de Silva et al., 2003, Cutillas et al., 2009). Ascariasis in humans is associated with deficiency of vitamin A, impaired cognition, stunting and asthma due to migration of worms toward the lungs (Lynn et al., 2021).

Control of STH is somehow difficult due to the high resistance of their egg/larvae to climatic factors and their ability to remain viable and infective in soil for years (Traversa *et al.*, 2014). However, the three key interventions crucial for long-lasting control of STH infection include anthelminthic drug treatment, improved sanitation and public health education (Tchuenté, 2011).

Soil-transmitted helminth infections in domestic pigs and dogs are reported worldwide (Agustina et al., 2021; Jia-Chi et al., 2016; Moskvina and Ermolenko, 2016; Agustina et al., 2022; Sharma et al., 2020), including in Africa (Chidumayo, 2018; Tumusiime et al., 2020; Nwafor et al., 2019). In the context of Tanzania, several studies have been conducted to establish the status of gastrointestinal helminth infection including STH in domestic pigs (Esrony et al., 1997; Mkupasi et al., 2010; Ngowi et al., 2004; Kabululu et al., 2015 and Nonga and Paulo, 2015) and dogs (Kidima, 2019; Makene et al., 1996 and Swai et al.,2010). Although several previous studies in Tanzania have reported the occurrence of gastrointestinal helminth infections in pigs and dogs, this study aimed to update the current status of STH of domesticated pigs and dogs useful to provide information that guides designing effective control measures.

Materials and Methods

The study area

This study was conducted in Mvomero District and Morogoro municipality of Morogoro region. The estimated population in the Morogoro region is 3,197,104 according to the Tanzania population and household census conducted in 2022. The study sites fall into the river valleys and basin agro-ecological zone with high temperatures (average of 30°C) and rainfall range between 900mm to 1400mm annually. The major economic activity in this region is agriculture including small-scale farming, livestock keeping, plantations and estates. The number of dogs in each study area was estimated to be 4624 (Morogoro municipality) and 5645 (Mvomero District). Mvomero District lies at latitude 06º 26' South and longitude 37° 32' East. It is bordered by the Tanga region (North), Pwani region (Northeast), Morogoro Rural District (East), Morogoro Urban (Southeast) and Kilosa District (West). It has a total area of 7,325 km² and has the largest water area (1,882 km²) in the region. Based on Tanzania's population and household census, the Mvomero District is divided into 4 divisions with 30 Wards and 130 villages. In this study, the samples were collected from two divisions

(Mzumbe and Mgeta) and four Wards (Nyandila, Mgeta, Lingali and Mlali). Morogoro municipality is found at latitude 06° 49' South and longitude 37° 39' East. It is located on the lower Uluguru Mountain slopes with a peak of about 500 to 600 above sea level. It has a total land area of 531 km². It has 29 Wards and 294 streets. Wards included in this study were Mazimbu, Kihonda Maghorofani, Kilakala, Bigwa and Kichangani (Figure 1).

Figure 1

Map of Tanzania (A) showing location of Morogoro region (B), Mvomero District (C) and Morogoro municipality (D)



Study design and selection of study sites

This was a cross-sectional study conducted on randomly sampled domestic pigs and dogs in the study sites. The selection of wards, villages, households and subsequently animals to be sampled was based on simple random method. At selected wards list of villages with communities domesticating domestic pigs and dogs was obtained from the ward offices and villages/streets were selected at random Similarly at selected villages/streets a list of households keeping domestic pigs and dogs was obtained from the local leaders and then participating households were selected at random

Sample size estimation

The sample size was determined by using the formula below according to (Sharma et al., 2020)

Sample size (n) = $((Z_{1-\alpha/2})^2(p)(q)) / d^2$

n stands for sample size, $Z_{1-\alpha/2}$ stands for critical value and a standard value for the corresponding level of confidence (at 95% CI or 5% level of significance is 1.96), p stands for prevalence that is presumed (0.5), q stands for 1-p and d stands for the margin of error (0.05).

Sample size for dogs according to the prevalence from Chidumayo, (2018); $n = ((1.96)^2 \times (0.24 \times 0.76)) / (0.05)^2 = 280$ faecal samples.

Sample size for domestic pigs according to Kabululu et al. (2015); $n = (1.96)^2 \times (0.175 \times 0.825)$ / (0.05)² = 222 faecal samples. Therefore, a total sample of 502 domestic pigs and dogs were sampled and examined for STHs infections.

Data collection

A data capture form was used to obtain information about the participating households (district, ward, the owner's name and contact) and the animals (age, sex, deworming status, housing condition or management system (indoor / semi-intensive / free roaming) and the cleaning of the housing facilities).

Prior to samples collection, the participating households were briefed about the purpose of the study and then faecal samples were collected from the randomly selected domestic pigs and dogs. During the faecal sample collection, the selected animals at household level were observed for some time until they defecated, then fresh faecal samples were collected from the ground using gloved hand and the sample put into a faecal container with lid. Each sample was labelled with the unique identification, age, sex, date and location of the household as previously described by Roepstorff and Nansen, (1998). The faecal samples were transported in a cool box with ice bags to Parasitology Laboratory at the College of Veterinary Medicine and Biomedical Sciences of Sokoine University of Agriculture and refrigerated at 4 - 8°C awaiting analysis.

Laboratory samples processing

The samples were processed by simple flotation method (Soulsby 1982), whereby 50 mL of flotation fluid was poured into a container and mixed with 3 g of feces using a tongue blade. The faecal suspension was filtered using a tea strainer and poured into a beaker. The test tube was placed into a beaker with the filtrate and waited for 10 minutes. The test tube was lifted from the beaker, transferred and ought to touch the surface of a microscopic slide for few seconds to allow the drop to run off. A cover slip was mounted on top of the microscope slide with the filtrate drop and examined under a microscope at 10x and 40x magnification as described by Roepstorff and Nansen (1998); Soulsby (1982) and Hansen and Perry (1990). Identification of the STH eggs were based on their size and shape as explained by Foreyt (1989).

Faecal samples found to have strongyle types of eggs were further subjected to faecal culture in order to ascertain the identity of the strongyle nematodes at genus level. Briefly, 10g of faecal sample were mixed with 4 g of sterile sawdust in a plastic container and distilled water was added to obtain the required consistency. The mixture was then left at room temperature for 7 days to allow eggs to hatch and larvae to develop of third stage. The third stage larvae were harvested by Baerman technique (Demelash et al., 2016). Briefly, harvesting was achieved as follows; a conical sedimentation flask was filled with tape water halfway, the culture wrapped in a double layer of gauze and fasten to a wood stick applicator. Thereafter, the culture was submerged in the tap water in the conical sedimentation flask and water was refilled 1cm to 3cm above the faeces and allowed to stay overnight (24 hours) for larvae to sediment. The following day the supernatant was discarded to obtain sediment containing the third stage larvae. Few drops from the sediment were transferred to a microscopic slide and a drop of iodine added. Thereafter, a coverslip was mounted on the slide that was examined at 10x magnification for identification of larvae using morphological features described by Abu – Elwafa et al., (2016) and Van Wyk et al., (2004).

Data management and statistical analysis

Data obtained for this study was entered using Microsoft Excel windows 10 and uploaded into STATA statistical software version 14 for cleaning and analysis. The prevalence (P) and chi square was computed. A *P*-value <0.05 was considered as statistically significant (Demeke *et al.*, 2021 and Salam *et al.*, 2009).

Ethics approval and consent to participateEthicalclearancenumber

SUA/DPRTC/R/186/Vol. IV - 26 was obtained from Sokoine University of Agriculture. All owners of animal participants were consented before sample and data collection.

Results

Proportion of STH infected animals in Morogoro Municipality and Mvomero District, Tanzania, based on location and animal information A total of 281 dogs were recruited, out of these 137 were from Morogoro Municipal Council and 144 from Mvomero District. Majority (54.1%) of the dogs were males with the median age of 2 years. A higher proportion of dogs from Morogoro Municipal Council (54.7%) were infected compared to those from Mvomero District (46.5%) (Table 1). Moreover, male, free roaming and dogs with no history of being dewormed were recorded to have more infection (Table 1). There was a positive association between STH infection with having no history of being dewormed whereby dewormed dogs were less likely to be detected with STH infection (pvalue = 0.002).

Table 1

Proportion of sampled dogs and pigs infected with soil transmitted helminths (STH) in Morogoro Municipality and Mvomero District, Tanzania, based on location and animal information

Variables		Dogs	Pigs
		Infected % (n/N)	Infected % (n/N)
Location	Morogoro Municipality	54.7% (75/137)	82.8% (101/122)
	Mvomero District	46.5% (67/144)	85.8% (101/115)
Sex	F	47.3% (61/129)	80.3% (106/132)
	Μ	53.3% (81/152)	91.4% (96/105)
Age	≤1 year	50% (37/74)	88.2% (157/178)
-	> 1 year	50.7% (105/207)	76.3% (45/59)
Deworming status	Yes	39.1% (43/110)	87% (127/146)
	No	57.9% (99/171)	82.4% (75/91)
Last deworming	Within 1 year	39.1% (43/110)	87% (127/146)
	Never	57.9% (99/171)	82.4% (75/91)
Management system	Free-roaming	53.3% (64/120)	-
	Semi-intensive	48.4% (78/161)	-
	Indoor	-	85.2% (202 / 237)

Of the 237 pigs examined, 122 were from Morogoro Municipality and 115 from Mvomero District Council. Majority of the pigs were females with a median age of 6 months. Location wise, pigs from Morogoro Municipal were most infected, while in terms of sex and age groups; male and pigs aged one year and below were more infected with STH species (Table 1). The difference in pig STH prevalence for age groups and sex was statistically significant with p-value of 0.025 and 0.016, respectively. However, dewormed pigs were most infected compared to those with no deworming history.

Prevalence of soil transmitted helminths in dogs Overall prevalence of STH in dogs was 50.53%, with *Ancylostoma* species being the most prevalent. Eggs encountered in faecal samples were those of *Ancylostoma* (47.33%), *Toxocara*

(12.46%) Trichuris (2.49%) and Strongyloides species (1.07%). Dogs in Mvomero District were more infected with Toxocara species while other detected STH species were more prevalent in Morogoro Municipal. Male dogs had relatively higher prevalence for almost all detected STH species with an exception Trichuris. Age group wise, adult's dogs were recorded to be more infected with Ancylostoma while younger dogs were observed to have more infected by Toxocara, Trichuris and Strongyloides. Dogs with deworming history within one year were recorded to have lowest infection compared with those that have never been dewormed in their lifetime. Lastly, dogs kept under semi intensive system had more infection in Strongyoloides while free roaming were having higher infection of Ancylostoma, Toxocara and Trichuris species (Table 2)

Table 2

Prevalence of STH Species in dogs from Morogoro Municipality and Mvomero District, Tanzania, based on location and animal information

STH species		Ancylostoma	Toxocara	Trichuris	Strongyloides
		%(n)	%(n)	%(n)	%(n)
Location	Morogoro Municipality (N=137)	52.6%(75)	10.9%(15)	3%(4)	2.9%(3)
	Mvomero (N=144)	42.4% (61)	13.9% (20)	2.1% (3)	0% (0)
Sex	F (N=129)	44.2%(57)	12.4% (16)	3.9%(5)	0% (0)
	M (N=152)	50%(76)	12.5% (19)	1.3% (2)	2% (3)
Age group	≤ 1 year (N=74)	45.9% (34)	14.9% (11)	2.7%(2)	2.7%(2)
001	> 1 year (N=207)	47.8% (99)	11.6% (24)	2.4% (5)	0.5% (1)
Deworming status	Yes (N=110)	35.5%(39)	6.5%(7)	0.9%(1)	0.9%(1)
status	No (N=171)	55%(94)	16.4%(28)	3.5%(6)	1.2%(2)
Last deworming	Within 1 year (N=110)	35.5%(39)	6.5%(7)	0.9%(1)	0.9%(1)
0	Never $(N=171)$	55%(94)	16.4%(28)	3.5%(6)	1.2%(2)
Management system	Free- roaming (N=120)	47.5% (57)	14.2%(17)	4.2% (5)	0% (0)
J -	Semi-intensive (N=161)	47.2% (76)	11.2% (18)	1.2%(2)	1.9% (3)

Prevalence of soil transmitted helminths in pigs In pigs, the overall prevalence of STH was 85.23% with the most prevalent species being *Oesophagostomum* (81.01%), followed by *Trichuris* (12.24) and *Ascaris* (5.49%). Male and pigs aged one year or below were more infected by all identified species of STH (*Oesphagostomum*, *Trichuris* and *Ascaris*). More pigs from Mvomero District were infected with two STH species compared to those from Morogoro, with the exception of *Trichuris species*. Pig with a history of deworming within one year had relatively more

infection by all STH reported in this study except *Ascaris* species (Table 3).

Table 3

Prevalence of STH Species in pigs from Morogoro Municipality and Mvomero District, Tanzania, based on location and animal information

STH species		Oesophagostomum %(n)	Trichuris %(n)	Ascaris %(n)
Location	Morogoro Mc(N=122)	77.9% (95)	23% (28)	4.9% (6)
	Mvomero (N=115)	84.3% (97)	0.9% (1)	6.1% (7)
Sex	F (N=132)	78.8% (104)	9.8% (13)	4.5%(6)
	M (N=105)	83.8% (88)	15.2% (16)	6.7% (7)
Age group	≤1 year (N=178)	82.6% (147)	16.3% (29)	5.6%(10)
	> 1 year (N=59)	76.3% (45)	0%(0)	5.1%(3)
Deworming status	Yes (N=146)	83.6% (122)	18.5%(27)	4.8% (7)
-	No (N=91)	76.9% (70)	2.2% (2)	6.6% (6)
Last deworming	Within 1 year	83.6% (122)	18.5%(27)	4.8% (7)
-	(N=146)			
	Never (N=91)	76.9% (70)	2.2% (2)	6.6% (6)
Management	Indoor (N=237)	81.01%(192)	12.2%(29)	5.5%(13)
system				

Mixed infection

This study observed that STH infection in pigs and dogs would occur either due to one or multiple types of parasite species (coinfection). Infection due to one type of STH species in pigs was recorded as Oesophagostomum 67.9% (n=161), Trichuris 2.1% (n=5) and Ascaris species 0.8% (n=2). However, the most common two STH species infection was observed for Oesophagostomum and Trichuris species 8.9% (n=21) followed by Oesophagostomum and Ascaris species 3.4% (n=8) and Trichuris and Ascaris species 0.4% (n=1). Moreover, for dogs single STH species infection occurred only for Ancylostoma 35.9 % (n=101) and Toxocara species 2.8% (n=8). However, two and three STH species coinfection was recorded as follows; Ancylostoma with Toxocara species 8.2% (n=23), Ancylostoma with Trichuris species 1.1% (n=3), Ancylostoma with Strongyloides species 1.1% (n=3), Toxocara with Trichuris 0.4% (n=1) and Ancylostoma, *Trichuris* and *Toxocara* 1.1% (n=3).

Discussion

Soil-transmitted helminths (STH) is one of the neglected tropical disease reported by WHO whose main source of infection is through contaminated soil. The infection is responsible for parasitic diseases in both human and animals with a great possibility for zoonosis. Ascaris suum, Ancylostoma and Toxocara species from pigs and dogs respectively, are reported to be of zoonotic importance (Nejsum et al., 2012; WHO, 2016). This study was designated to investigate the prevalence of STH among domestic pigs and dogs to give information that would help to prioritize prevention and control interventions of STH infections. This would help to improvement animal health and minimize risk of zoonotic parasite to be transmitted to human beings. This study revealed that STH are major gastrointestinal parasite infecting pigs and dogs.

The observed STH prevalence in pigs (85.23%) was higher compared to the previous studies conducted in Tanzania which recorded for gastrointestinal helminths prevalence of 53% and

63.7% (Esrony et al., 1997; Kabululu et al., 2015). Higher STH prevalence than previous studies done in Tanzania may be due increased pig population thus it became difficult to apply good animal husbandry practice leading to increased parasite circulation in the ecosystem. Moreover, climatic change may have favored the survival of the infective eggs and larva in the environment. Similar findings to this study have been reported in Rwanda (84.6%) and Kenya (84.2%) onto which STH were the most prevalent gastrointestinal parasites in pigs with Strongyletype helminths being the most prevalent egg species (Kagira et al., 2012; Tumusiime et al., 2020). This similarity may be because of the traditional rearing system adopted in these countries having poor management systems, housing, hygiene and feeding along with lack of regular deworming of pigs giving room to continuous infestations and re-infection (Nganga et al., 2008; Roepstorff et al., 2011). Generally, High prevalence STH infection in African countries may be due to lack of effective disease prevention and control strategies (Tamboura et al., 2006).

Furthermore, this study has reported the occurrence of three STH species infecting pigs that include Oesphagostomum species, Trichuris suis and Ascaris suum. Similar species have been reported by previous studies in Tanzania and elsewhere (Gabon, South Africa and Kenya) (Esrony et al., 1997; Kabululu et al., 2015; Kagira et al., 2012; Maganga et al., 2019; Nwafor et al., 2019). Nevertheless, there was a significant association of sex and age with STH infestations. Younger pigs were the most infected age group with all three detected STH species while infection rate was low in adult. Similar to other studies, this was associated with stronger immune memory acquired by adult pigs after several prior infestations that enable them to expel immature parasite in the small intestine before they are able cause morbidity and/or lay eggs (Kagira et al., 2008; Roepstorff and Nansen 1994; Tamboura et al., 2006).

Additionally, this study reported that male pigs were more infected with all three detected STH species than females which was in agreement with Sowemimo *et al.*, (2012), a study done in Southwest Nigeria. Contrary, other studies

reported a non-significant effect of sex to gastrointestinal helminths infestation and female pigs had higher prevalence prevalence (Maganga et al., 2019; Sharma et al., 2020; Tamboura et al., 2006). High rooting behavior of male pigs that lead to increased access to infected soil was associated with higher prevalence for male pigs in comparison to females (Forrester et al., 1982). Interestingly in this study, pigs with history of deworming within one year had relatively higher infection especially for Oesphagostomum species and Trichuris suis which is different from other studies (Kabululu et al., 2015; Tamboura et al., 2006). This study relied on a self-reported antihelminthic treatment practiced by farmers. Farmers might be missing the correct timing to administer antiparasitic drug and might be administering less effective drugs in the incorrect dosage (Roesel et al., 2016). Moreover, without effective STH control strategies, there is a high chance of reinfection especially in the already contaminated environment (Nganga et al., 2008).

Moreover, in this study dogs were presented with an overall STH prevalence of 50.53% which was lower in comparison to previous studies within Tanzania (Makene et al., 1996. Kidima, 2019). This may be due to deworming of animals in the present study as Kidima, 2019 report was based on stray dogs. Moreover, the present advancement in control strategies (proper management system and health education) from 1996 may have influenced the low prevalence in the present findings; however, eco - climatic conditions that could occurred overtime may have played part (Jia-Chi et al., 2016). Nevertheless the obtained prevalence in this study was higher compared to previous study in Bali, Indonesia that reported a prevalence of 38% (Agustina et al., 2021). The inconsistence of the prevalence in these countries might be due to differences in soil nature, climatic conditions and the altitude of the study sites (Moskvina and Ermolenko, 2016; Jia-Chi et al., 2016). The STH detected from dogs in the present study were Ancylostoma, Toxocara, Trichuris and Strongyloides species, with the Ancylostoma species being most prevalent in all age groups. The later observation concurs with previous studies in Tanzania and elsewhere in the world (Makene et al., 1996; Kidima, 2019). Contrary to previous studies in Tanzania (Makene et al., 1996; Kidima, 2019),

present study reports the occurrence of Trichuris species in dogs for the first time. Male dogs had relatively higher prevalence of the STH infestation even though sex was not reported to significantly influence STH prevalence of dogs in Similarly, study done earlier in this study. Tanzania reported that sex had no effect on infection prevalence (Makene et al., 1996). However, it was reported in Nigeria that male significantly had higher infection dogs prevalence than females (Anosike et al., 2004). The later results were associated with the fact that male dogs tend to be more free-ranging when are compared to female. Free roaming dogs has higher chances to come into contact with contaminated soil and hence become infected by many types of STH species (Jia-Chi et al., 2016). This explains why free-roaming dogs in this study had more STH infections compared to those kept under semi-intensive management system.

Nevertheless, age group wise the infection rate was comparable for almost all STH detected in this study indicating that age did not influence the STH prevalence. However, these results were contrasting to a study done earlier in Morogoro which pointed out that age was significantly associated with gastrointestinal helminths prevalence where higher prevalence were observer for puppies (Makene et al., 1996). The later attributed their results to transplacental and transmammary infection transmission routes in puppies and/or continued contamination of the environmental especially by asymptomatic adult dogs (Makene et al., 1996). Conversely, adult had relatively higher prevalence of Ancylostoma species than young dogs which might be because this was the most prevalent species reported for this study. These results are also consistent with study of (Kidima, 2019) done in Rukwa, Katavi and Arusha and (Swai et al., 2010) done in Arusha, Tanzania who had reported the high prevalence of hookworm. High prevalence of hookworm infection in dog was associated with increased availability of hookworm paratenic hosts, vertical transmission in utero and puppies acquisition of hookworm through nursing (Kidima, 2019).

Dogs with deworming history within one year were recorded to significantly have lowest STH

infection when compared with those that have never been dewormed in their lifetime. Deworming has been associated with reduced worm burden in animals which might be the explanation of low infection rate in dewormed animals that participated in this study. Moreover, regular deworming was suggested to reduce the effect of sex on helminths infection susceptibility (Chidumayo, 2018).

Nonetheless, hookworms and roundworms of pigs and dogs are the most important helminths imposing risk to animals and human health. The present findings reported the occurrence of zoonotic STHs in dogs (Ancylostoma and Toxocara species) and pigs (Ascaris species). It was observed that zoonotic species were the most prevalent in dogs while in pigs was the least detected species. Ancylostomiasis has been associated with chronic hemorrhagic and microcvtic hypochromic anemia. Besides, Toxocara species may be acquired direct to human being even through pets' hair containing infective eggs. Toxocariasis in human can cause ocular and visceral toxocariasis that is more prevalent in children and adult, respectively (Moskvina & Ermolenko, 2016).

Conclusion

The study findings indicate that STHs species are prevalent among dogs and pigs kept in the study sites. Owners of the pigs and dogs are prudently advised to undertake appropriate control measures against the STH infection to improve animal health and production where applicable. Furthermore, the appearance of zoonotic parasites (*Ancylostoma* species, *Toxocara* species and *Ascaris suum*) calls for one health interventions to control the parasites in order to safeguard the public health.

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References

- Abu Elwafa, S. A., Al Kappany, Y. M. & El -Alfy, E. N. (2016). Identification of nematodes third stage larvae of ruminant animals. *Egyptian Veterinary Medical Society* of Parasitolody Journal. (12) 60-73
- Anosike, J.C., Nwoke, B.E., Ukaga, C.N., Madu, N.G., & Dozie, I.N., (2004). Aspects of intestinal helminth parasites of dogs in World bank-assisted Housing Estate, New Owerri, Nigeria. African Journal of Applied Zoology & Environmental Biology; 6, 25–29.
- Agustina, K. K., Anthara, M. S., Sibang, N. A. A. N., Wiguna, W. A. R., Apramada, J. K., Gunawan, W. N. F., Oka, I. B. M., Subrata, M., & Besung, N. K. (2021). Prevalence and distribution of soil-transmitted helminth infection in free-roaming dogs in Bali Province, Indonesia. *Veterinary world*, 14(2), 446–451. https://doi.org/10.14202/vetworld.2021.4 46-451
- Agustina, K. K., Wirawan, I. M. A., Sudarmaja, I. M., Subrata M. & Dharmawan, N. S. (2022). The first report on the prevalence of soiltransmitted helminth infections and associated risk factors among traditional pig farmers in Bali Province, Indonesia, *Veterinary World*, 15(5): 1154-1162
- Chidumayo N. N. (2018). Epidemiology of canine gastrointestinal helminths in sub-Saharan Africa. *Parasites & vectors*, 11(1), 100. https://doi.org/10.1186/s13071-018-2688-9
- Crompton, D.W.T. (2000) The public health importance of hookworm disease. *Parasitology*; 121(S1): S39-S50.
- Cutillas, C., Callejon, R., de Rojas M., Tewes, B., Ubeda, J.M., Ariza, C. & Guevara, D.C. (2009). Trichuris suis and Trichuris trichiura are different nematode species. *Acta Tropica*; 111, 299–307
- de Silva, N.R., Brooker, S., Hotez, P.J., Montresor, A., Engels, D. & Savioli, L. (2003). Soiltransmitted helminth infections: updating

the global picture. *Trends Parasitol.* 19, 547–551

- Demeke, G., Fenta, A., & Dilnessa, T. (2021). Evaluation Wet of Mount and Concentration Techniques of Stool Examination for Intestinal Parasites Debre Identification Markos at Comprehensive Specialized Hospital, Ethiopia. Infection and drug resistance, 14, 1357-1362. https://doi.org/10.2147/IDR.S307683
- Demelash, K., Abebaw, M., Negash, A., Alene, B., Zemene, M., & Tilahun, M. (2016). A Review on Diagnostic Techniques in Veterinary Helminthlogy. *Natural Science*, 14(7), 109–118. <u>https://doi.org/10.7537/marsnsj140716.15</u> <u>.Key</u>
- Esrony, K., Kambarage, D. M., Mtambo, M. M., Muhairwa, A. P., & Kusiluka, L. J. (1997). Helminthosis in local and cross-bred pigs in the Morogoro region of Tanzania. *Preventive veterinary medicine*, 32(1-2), 41–46. https://doi.org/10.1016/s0167-5877(97)00011-1
- Foreyt W. (1989) Diagnostic Parasitology. Veterinary Clinics of North America: Small Animal Practice, **19**(5), 979–1000. https://doi.org/10.1016/S0195-5616 (89)50107-4
- Forrester, D.J., Porter, J.H., Belden, R.C. & Frankenberger, W.B. (1982): Lungworms of feral swine in Florida. *Journal of American Veterinary Medical Association*; 181: 1278-1280
- Hansen J. and Perry B., (1990). The Epidemiology, Diagnosis, and Control of Helminth Parasites of Ruminants, *International laboratory for Reseach on Animal Diseases*, Nairobi, First edition.
- Jia-Chi, C., Abdullah, N. A., Shukor, N., Jaturas, N., Richard, R. L., Majid, M. A. A., ... & Nissapatorn, V. (2016). Asian Pacific Journal of Tropical Disease Soil transmitted helminths in animals – how is it possible for

human transmission ? *Asian Pacific Journal of Tropical Disease*, 6(11), 859–863. https://doi.org/10.1016/S2222-1808(16)61146-5

- Kabululu, M. L., Ngowi, H. a, Kimera, S. I., Lekule, F. P., Kimbi, E. C., & Johansen, M. V. (2015). Veterinary Parasitology Risk factors for prevalence of pig parasitoses in Mbeya Region, Tanzania. *Veterinary Parasitology*, 212(3-4), 460–464. https://doi.org/10.1016/j.vetpar.2015.08.0 06
- Kagira, J. M., Kanyari, P. N., Githigia, S. M. and Maingi, N. (2012). Risk factors associated with occurrence of nematodes in free range domestic pigs in Busia District , Kenya. *Tropical animal health and production*, 44(3) 657–664. https://doi.org/10.1007/s11250-011-9951-9
- Kagira, J.M., Kanyari, P.W.N., Waruiru, R.M. & Munyua, W.K. (2008). Relationship between the prevalence of gastrointestinal nematode infections and management practises in pig herds in Thika District, Kenya. *Livestock Research for Rural Development*; 20 (10)
- Kidima, W. (2019). Prevalence of Zoonotic Parasites in Stray Dogs in Rural Communities. *African Journals online*; 45(1), 93–100.
- Lynn, M. K., Morrissey, J. A., & Conserve, D. F. (2021). Soil-Transmitted Helminths in the USA: a Review of Five Common Parasites and Future Directions for Avenues of Enhanced Epidemiologic Inquiry. *Current tropical medicine reports*, 8(1), 32–42. https://doi.org/10.1007/s40475-020-00221-2
- Maganga, G. D., Kombila, L. B., Boundenga, L., Kinga, I. C. M., Obame-Nkoghe, J., Tchoffo, H., Gbati, O. B., & Awah-Ndukum, J. (2019). Diversity and prevalence of gastrointestinal parasites in farmed pigs in Southeast Gabon, Central Africa. *Veterinary world*, 12(12), 1888–1896. https://doi.org/10.14202/vetworld.2019.1 888-1896
- Makene, V. W., Muhairwa, A. P., & Kambarage,

D. M. (1996). Prevalence of Canine Gastrointestinal Parasites in Morogoro , Tanzania, (April 2015). *Journal of Applied Animal Research;* 10:2, 149-153. https://doi.org/10.1080/09712119.1996.97 06142

- Massetti, L., Colella, V., Zendejas, P. A., Ngnguyen, D., Harriott, L., Marwedel, L. & Traub, R. J. (2020). High-throughput multiplex qPCRs for the surveillance of zoonotic species of canine hookworms. *PLOS neglected tropical diseases*. 14(6): e0008392. https://doi.org/10.1371/journal.pntd.0008 392
- Mkupasi, E. M., Ngowi, H. A., & Nonga, H. E. (2010). Prevalence of extra-intestinal porcine helminth infections and assessment of sanitary conditions of pig slaughter slabs in Dar es Salaam city, Tanzania. *Tropical animal health and production*, 43(2), 417–423. https://doi.org/10.1007/s11250-010-9708x
- Moskvina, T. & Ermolenko, A. (2016). Helminth infections in domestic dogs from Russia. *Veterinary World*, (EISSN: 2231-0916).
- Nejsum, P., Betson, M., Bendall, R.P., Thamsborg, S.M. & Stothard, J.R. (2012). Assessing the zoonotic potential of *Ascaris suum* and *Trichuris suis*: looking to the future from an analysis of the past. *Journal of Helminthology*; 86;148–155. doi:10.1017/S0022149X12000193
- Nganga, C. J., Karanja, D. N., & Mutune, M. N. (2008). The prevalence of gastrointestinal helminth infections in pigs in Kenya. *Tropical animal health and production*, 40(5), 331–334. https://doi.org/10.1007/s11250-007-9112-3
- Ngowi, H. A., Kassuku, A. A., Maeda, G. E. M., Boa, M. E. & Willingham, A. L. (2004). A Slaughter Slab Survey for Extra-Intestinal Porcine Helminth Infections in Northern Tanzania. *Tropical animal health and production*, 36(4); 335–340. https://doi.org/10.1023/b:trop.000002666 3.07862.2a

Nonga, H. E. & Paulo, N. (2015). Prevalence and

intensity of gastrointestinal parasites in slaughter domestic pigs at Sanawari slaughter slab in Arusha , Tanzania. *Livestock Research for Rural Development*; 27 (1).

- Nwafor, I. C., Roberts, H. & Fourie, P. (2019). Prevalence of gastrointestinal helminths and parasites in smallholder domestic pigs reared in the central Free State Province. *Onderstepoort Journal of Veterinary Research*. 11;86(1):e1-e8. doi: 10.4102/ojvr.v86i1.1687. PMID: 31038321; PMCID: PMC6495001.1–8.
- Roepstorff, A. & Nansen, P. (1994) Epidemiology and control of helminth infection in pigs under intensive and non-intensive production systems. *Veterinary Parasitology*, 54, 69–85.
- Roepstorff, A., & Nansen, P. (1998). Epidemiology, Diagnosis And Control Of Helminth Parasites Of Swine. Food and Agriculture Organization Of The United Nations, ISBN 92-5-104220-9
- Roepstorff, A., Mejer, H., Nejsum, P. & Thamsborg, S.M. (2011). Helminth parasites in pigs: New challenges in pig production and current research highlights. *Veterinary Parasitology* 180; 72– 8
- Roesel, K., Dohoo, I., Baumann, M., Dione, M., Grace, D. & Clausen, P. (2016) Prevalence and risk factors for gastrointestinal parasites in small-scale pig enterprises in Central and Eastern Uganda, *Parasitology Research*. DOI:10.1007/s00436-016-5296-7
- Salam, M. M., Maqbool, A., Naureen, A., & Lateef, M. (2009). Comparison of different diagnostic techniques against Fasciolosis in Buffaloes. *Veterinary World*; 2(4); 129–132.
- Sharma, D., Singh, N. K., Singh, H., & Rath, S. S. (2020). Copro-prevalence and risk factor assessment of gastrointestinal parasitism in Indian domestic pigs. <u>*Helminthologia*</u>; 57(1); 28–36. https://doi.org/10.2478/helm
- Soulsby, E. J. (1982). Helminth, Arthropods and Protozoa of Domesticated Animals. *Bailliere Tindall, London, seventh ed*, 212–252.
- Sowemimo, A.O., Asaolu, S.O., Adegok, F.O. & Ayanniyi, O.O., (2012). Epidemiological

survey of gastro-intestinal parasites of pigs in Ibadan, Southwest Nigeria. *Journal of Public Health and Epidemiology*, 4; 294–298. https://doi.org/10.5897/JPHE12.042

- Strube, C., Heuer, L., & Janecek, E. (2013). Toxocara spp. infections in paratenic hosts. *Veterinary Parasitol;* (193), 375–389. Retrieved from https://doi.org/10.1016/j.vetpar.2012.12.0 33
- Swai, E.S., Kaaya, E.J., Mshanga, D.A. and Mbise E.W. (2010) A Survey on Gastro-Intestinal Parasites of Non-Descript Dogs in and Around Arusha Municipality, Tanzania. *International Journal of Animal and Veterinary Advance;* 3(2): 63-67
- Tamboura, H.H., Banga-Mboko, H., Maes, D., Youssao, A.K., Traore, A., Bayala, B., & Dembele, M.A. (2006). Prevalence of common gastrointestinal nematode parasites in scavenging pigs of different ages and sexes in eastern centre province, Burkina Faso," Onderstepoort Journal of Veterinary Research 73(1); 53-60.
 - Tchuenté, L. A. T. (2011). Acta Tropica Control of soil-transmitted helminths in sub-Saharan Africa : Diagnosis , drug efficacy concerns and challenges. *Acta Tropica*; 120(1); 4–11. https://doi.org/10.1016/j.actatropica.2010 .07.001
 - Traub, R. J., Zendejas-Heredia, P. A., Massetti, L., & Colella, V. (2021). Zoonotic hookworms of dogs and cats – lessons from the past to inform current knowledge and future directions of research. *International Journal of Parasitol.*, (51); 1233–1241. Retrieved from https://doi.org/10.1016/j.ijpara.2021.10.00 5
 - Traversa, D. (2012). Pet roundworms and hookworms: A continuing need for global worming. *Parasite and Vectors*; (5(91)), 1–19.
 - Traversa, D., Frangipane, A., Cesare, A. Di, Torre,
 F. La, Drake, J., & Pietrobelli, M. (2014).
 Environmental contamination by canine geohelminths. *Parasites Vectors*; 7(67); 1–9. https://doi.org/10.1186/1756-3305-7-67

Tumusiime, M., Ntampaka, P., Niragire, F.,

Sindikubwabo, T. & Habineza, F. (2020). Prevalence of Swine Gastrointestinal Parasites in Nyagatare District , Rwanda, *Journal of Parasitology Research;* 2020, 8814136.

https://doi.org/10.1155/2020/8814136

- Van Wyk, J.A., Cabaret J., & Michael L.M., 2004. Morphological identification of nematode larvae of small ruminants and cattle simplified. *Veterinary parasitology;* (199), 277-306.
- Walker, N. I., Croese, J., Clouston, A. D., Parry, M., Loukas, A., & Prociv, P. (1995).
 Eosinophilic enteritis in Northeastern Australia: pathology, association with Ancylostoma caninum, and implications.

The American journal of surgical pathology; (19), 328–337. Retrieved from https://doi.org/10.1371/journal.pntd.0009 395

- Walsh, M. G., & Haseeb, M. A. (2014). Toxocariasis and lung function: Relevance of a neglected infection in an urban landscape. *Acta Parasitol.* (59), 126–131.
- World Health Organization (WHO) (2016). Zoonoses and the Human-Animal-Ecosystems Interface. *World Health Organization, Geneva*.
- Yacob, H. T., Ayele, T., Fikru, R. & Basu, A. K. (2007). Gastrointestinal nematodes in dogs from Debre Zeit, Ethiopia. *Vet. Parasitol.*, 148(2), 144–148.